

REVIEW ARTICLE

The Diversity of Bacteriocin and Its Antiviral Potential: An Overview

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ABSTRACT

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The viral resistances to the known antiviral agent have raised the need for developing alternatives antiviral agents. one of the promising alternatives are bacteriocins. Bacteriocin are natural peptides ribosomal synthesized by gram negative and gram positive bacteria. Bacteriocins are secreted as a method for the producer to compete in the environment. Bacteriocin gene cluster can be found on plasmid or chromosome. Bacteriocins have presented wide range of activity against bacteria, fungi, virus and cancer cells and have a high margin of safety which encouraged its use in food and pharmaceutical applications. Nisin is one example of bacteriocins approved by FDA for use as food preservative. Bacteriocin nature variation is one of its unique qualities. Bacteriocin varies in physical, chemical, molecular and genetic nature. This variation have been studied and recoded over the years. In this review we discuss and summarized the diversity of bacteriocins and their different mechanism of actions against virus.

INTRODUCTION

The current pandemic¹ and the continuous resistance of antiviral agents have raised the need for antiviral agents² meanwhile bacteriocins have been suggested as an alternative antiviral agents³. Bacteriocins are ribosomal synthesized peptides produced by gram negative and gram positive bacteria⁴. It is considered a golden weaponry arsenal against different pathogen⁵ because of its high safety margin⁶ and the diversity of its structures and characters⁷. The first bacteriocin was discovered in the year 1925 by Gratia from *Escherichia coli* and later named as colicin⁸. Since then large number of bacteriocins have been identified from a diverse group of bacterial strains. bacteriocins were also successfully applied in the food industry as bio preservative⁹ e.g., Nisin which have been approved by FDA¹⁰. Moreover several applications have been reported for bacteriocins such as antimicrobial¹¹ and anticancer¹² which encouraged the attempts to use bacteriocins in pharmaceutical industry¹³. In this review we discuss the diversity of bacteriocins and their different mechanism of actions against virus.

Bacteriocin from Gram positive bacteria

Lantibiotics:

Lantibiotics are <5kda peptides that can endure heat and PH¹⁴. They are called Lantibiotics because they have lanthionine, methyl-lanthionine and unsaturated amino

acids residue which form rings¹⁵. Lantibiotics have been classified as class I of gram positive bacteria and can be sub-categorized to class Ia and class Ib. Class Ia is flexible elongated structure that acts by binding of N-terminal to lipid II which inhibits peptidoglycan and C-terminal form pores in the cell membrane. On the other hand class b are negative charge inflexible globular structures that act by inhibiting cell enzymes¹⁶.

Non -lantibiotics: Class II, III and IV.

Class II are small heat stable that is not post transitionally modified except for disulfide bond¹⁷. This class have four sub classes IIa, IIb, IIc and IId. Class IIa is a linear Pediocin like bacteriocin with a distinct amino acid sequence that is active against *Listeria monocytogenes*. Class IIb is two peptides that act together for activity class IIC contains cysteine and are called thiolbiotics and cystibiotics and lastly class IId contain the class II bacteriocin that doesn't belong to the other three sub classes¹⁸. The class II are cationic that act on target microorganisms by permeabilization of the membrane¹⁹. Class III are heat sensitive >30kda protein and class IV have a lipid or/and carbohydrate part²⁰.

Bacteriocins from gram negative bacteria.

Microcins:

Microcins are < 10 kda peptides that can endure drastic PH and temperature also protease enzyme²¹. They are entitled as microcins due to its distinct low

molecular weight ²² Microcins have been grouped to class I which are post-translationally modified and have < 5 kDa molecular weight and class II which are seldom modified post-translationally with molecular weight between 5 and 10 KDa²³. Microcins have been found to exert its activity against target microorganism by either pore formation in cell membrane²⁴ or inhibiting enzymes such as DNA gyrase ²⁵.

Colicins:

Colicins are >10kDa high molecular weight protein and heat sensitive produced by *E.coli* as an SOS reaction which is a DNA repair system that allows DNA replication to bypass errors in the DNA induced by the environment²⁶. Colicins have been grouped to group A which require Tol protein to cross the cell outer membrane²⁷ and group B which requires TonB protein to cross the cell outer membrane²⁸.

Classification of bacteriocins

The diversity of bacteriocins and its producers have hindered its grouping and classifications. Many approaches have been proposed to classify and group bacteriocins ²⁹⁻³⁰. One of the most distinct approaches was suggested by Cotter et al ¹¹ which is simple and

includes both gram positive and negative bacteria. This approach removed large protein such as colicins to include peptides only and used post-translationally modified peptides (RiPPs) nomenclature³¹. Cotter et al.¹¹ classified bacteriocins to Class I (modified post-translationally bacteriocins) and class II (Unmodified bacteriocins). Class I was further sub-categorized to lantibiotics, proteusins, cyanobactins, thiopeptides, sactibiotics, bottromycins, glycocins, prenylated, anacyclamide-like cyanobactins, patellamide-like cyanobactins, lasso peptides, linaridins, linear azole and modified microcins that are not related to the above subgroups and class II was further sub-categorized to IIa, IIb, IIc, II d and IIe. Recently Soltani et al³² suggested an update to this classification by grouping class I as post-translationally modified peptides < 5 KDa with related enzymes embedded in its gene cluster while class II 4-6 KDa unmodified peptides that may or may not have a disulfide bridge. This update signifies class I with higher stability than class II despite the disulfide link. The summary and comparison of Cotter et al ¹¹ and Soltani et al.³² classification of bacteriocins is presented in (table 1).

Table 1: Summary and comparison of Cotter et al¹¹ and Soltani et al³² classification of bacteriocins.

Class I: post-translationally modified bacteriocins.					
Cotter et al ¹¹			Soltani et al ³²		
GROUP	properties	Examples	GROUP	properties	examples
Lantibiotics	Lanthionine bridge	Nisin, actagardine, mersacidin, planosporicin, and mutacin 1140	Lantibiotics	Lanthionine bridge and two peptides lantibiotics	Nisin, actagardine, mersacidin, planosporicin, and mutacin 1140 Two peptides :Lacticin 3147 and Haloduracin
Sactibiotics	sulphur- α -carbon link	Ruminococcin C, thuricin CD, Subtilosin A.	Sactibiotics	sulphur- α -carbon link	Ruminococcin C, thuricin CD, Subtilosin A.
Linaridins	Linear peptides with dehydrated amino acid	Cypemycin	Linaridins	Linear peptides with dehydrated amino acid	Cypemycin
Thiopeptides	Heterocyclic, pyridine, piperidine, and dihydropyridine	Thiostrepton	Thiopeptides	Heterocyclic, pyridine, piperidine, and dihydropyridine	Thiostrepton
Glycocins	Glycopeptides with S-link	Sublancin 168	Glycocins	Glycopeptides with S-link	Sublancin 168
Linear azole	Peptide linear that have heterocyclic oxazole and thiazole	Microcin B17	Linear azole	Peptide linear that have heterocyclic oxazole and thiazole	Microcin B17
Bottromycins	Macrocyclic with amidine moiety, decarboxylated thiazole and methylated amino acids	Bottromycin A2	Bottromycins	Macrocyclic with amidine moiety, decarboxylated thiazole and methylated amino acids	Bottromycin A2
Cyanobactins	Macrocyclic peptides with heterocycles and	Patellamide A	Cyanobactins	Macrocyclic peptides with heterocycles and	Patellamide A

	may or may not have a prenylated amino acid.			may or may not have a prenylated amino acid.	
Lasso peptides	Possess lasso form.	Microcin J25	Lasso peptides	Possess lasso form.	Microcin J25
MccC7-C51-type bacteriocins	have aspartic acid with carboxy-terminal	Microcin C7 to C51	Nucleotide peptides	have a nucleotide fragment	Microcin C
Proteusins	Have several methylation ,hydroxylation and epimerization	Polytheonamide A	Siderophore peptides	Possess siderophore type non ribosomal modified linked to C terminal serine containing part	Microcins H47, Microcin E492
			Circular peptides	Single non modified peptides Cyclized by N to c linkage	Gassericin A , Garvicin ,Enterocin AS-48
Class II: Non- modified bacteriocins.					
Cotter et al¹¹			Soltani et al³²		
GROUP	properties	examples	GROUP	properties	examples
Peptides IIa	Have YGNGV sequence	enterocin CRL35, Pediocin PA-1, carnobacteriocin BM1	single Pediocin-like peptides	Have YGNGV sequence	enterocin CRL35, Pediocin PA-1, carnobacteriocin BM1
Peptides IIb	Two peptides are necessary for activity	Lactacin F	Two-peptides	Two non-modified or more peptides are necessary for activity	Lactacin F
Peptides IIc	Peptides in cycle form	Enterocin AS-48			
Peptides IIId	non-pediocin like single peptides	Lactococcin A, Epidermicin NI01, Microcin V	single Unmodified peptides	non-pediocin like single peptides	Lactococcin A, Epidermicin NI01, Microcin V
Peptides IIE	Possess siderophore type non ribosomal modified linked to C terminal serine containing part	Microcin E492			

Bacteriocins oppose virus:

The bacteriocins antiviral activity was reported for enterocin CRL35 produced by *Enterococcus faecium* CRL35³³. It was found that enterocin CRL35 inhibited *Herpes Simplex* (HSV) type 1 and 2 but wasn't virucidal and suggested that enterocin CRL35 acted on viral intracellular multiplication which was further investigated by Wachsmann et al³⁴ whom concluded that enterocin CRL35 acted on glycoprotein synthesis on the viral replication but didn't affect its uptake. Similarly another report showed that enterocin ST4V produced by *Enterococcus mundtii* ST4V inhibited HSV type 1 and 2 by 99.9%³⁵ while polio virus was inhibited by only 50% and measles virus by 95% ,however Cavicchioli et al³⁶ reported that *Enterococcus durans* Gen12 had antiviral activity against polio virus of 93.7% after adsorption and 27.9% against HSV type 1 before adsorption. Additionally enterocin ST5Ha displayed activity against HSV type 1³⁷ and enterocin B displayed activity against H3N2, H1N1³⁸. Although these report mechanisms are still unknown, it is

interestingly to note that most are considered class II bacteriocins. Another mechanism of action was reported by Féris et al³⁹ for Labyrinthopeptin A1 which is a lantibiotics against HIV and HSV. Labyrinthopeptin A1 inhibits HIV transmission between T-cells by inhibiting entry also by acting on the virus envelope but not the receptor. on the other hand Torres et al⁴⁰ suggested that Subtilosin A at a concentration lower than virucidal doesn't act before HSV type 1 and 2 viral protein synthesis which impose that Subtilosin A act on either assembly or the release this was in agreement with Quintana et al⁴¹ findings. The binding of Duramycin to phosphatidylethanolamine in zika virus envelope hindered TIM1 receptor and lowered infection in placental cells and explants⁴². Similar mechanism was observed for Ebola, West Nile and dengue viruses⁴³. Hepatitis C virus entry was hindered by Micrococcin P1 also cell to cell transmtion without affecting viral particles secretion⁴⁴. Bacteriocins with antiviral activity are compared and summarized in (table 2).

Table 2: Bacteriocin that have exhibited activity against virus.

Bacteriocin	Properties of bacteriocins	Producer microorganism	Virus active against	Suggested mechanism for antiviral activity	Reference
Enterocin CRL35	ClassII/ pediocin-like bacteriocin	<i>Enterococcus faecium CRL35</i>	HSV type 1 HSV type 2	Late stage glycoprotein synthesis on the viral replication	33,34
Enterocin B	Chemically synthesized	<i>Enterococcus faecium L3</i>	H3N2 H1N1	Not determined	38
Enterocin ST4V	Non-glycosylated 3950 Da peptide	<i>Enterococcus mundtii ST4V</i>	HSV type 1 HSV type 2 polio virus measles virus	Not determined	35
Enterocin ST5H	ClassII/ pediocin-like bacteriocin	<i>Enterococcus faecium ST5Ha</i>	HSV type 1	Not determined	37
Subtilisin A	Sactibiotics	<i>Bacillus subtilis KATMIRA 1933</i>	HSV type 1 HSV type 2	act on assembly or the release	40,41
Labyrinthopeptin A1	lantibiotics	<i>Actinomadura namibiensis DSM 6313</i>	HSV type 1 HIV	inhibiting entry also by acting on the virus envelope but not the receptor	39
Duramycin	cyclic 19-aa peptide	<i>Streptomyces cinnamoneus</i>	Zika virus West Nile virus Dengue virus Ebola virus	inhibit TIM1 receptor	42,43
Micrococin P1	macrocyclic peptide	<i>Staphylococcus equorum WS2733</i>	Hepatitis C virus	Acted on virus cell entry without affecting the secretion of viral particales	44
Bacteriocin in cell free supernatant	Bacteriocin like substance	<i>Lactobacillus delbrueckii</i>	Influenza virus H7N7 H7N1	Decreased the Expression of hemag- glutinin ,viral glycoproteins,neuraminidase, and nucleoprotein on the surface of infected cells,and hemagglutinin production and virus yield,	45
Semi-purified bacteriocins		<i>Lactococcus lactis GLc03 and GLc05, E. durans GEn09, GEn12, GEn14 and GEn17</i>	HSV type 1 polio virus	Acted after adsorption of polio virus and may have acted on HSV type 1 envelope or affected its binding to the cell receptor	36

This manuscript has not been previously published and is not under consideration in the same or substantially similar form in any other reviewed media. I have contributed sufficiently to the project to be included as author. To the best of my knowledge, no conflict of interest, financial or others exist. All authors have participated in the concept and design, analysis, and interpretation of data, drafting and revising of the manuscript, and that they have approved the manuscript as submitted.

REFERENCES

- Xie Y, Wang Z, Liao H, Marley G, Wu D, Tang W. Epidemiologic, clinical, and laboratory findings of the COVID-19 in the current pandemic: systematic review and meta-analysis. *BMC Infect Dis.* 2020;20;1:1-12.
- Cento V, Chevaliez S, Perno CF. Resistance to direct-acting antiviral agents: clinical utility and significance. *Curr Opin HIV AIDS.* 2015;10;5:381-389.
- Al Kassaa I, Hober D, Hamze M, Chihib NE, Drider D. Antiviral Potential of Lactic Acid Bacteria and Their Bacteriocins. *Probiotics Antimicrob Proteins.* 2014;6, 3-4:177-185.
- Da Costa RJ, Voloski FLS, Mondadori RG, Duval EH, Fiorentini ÂM. Preservation of meat products with bacteriocins produced by lactic acid bacteria isolated from meat. *J Food Qual.* 2019;1;1-12.
- Negash AW, Tsehai BA. Current applications of

- Bacteriocin. *Int J Microbiol.* 2020;1;1-7.
6. Gautam N, Sharma N. Bacteriocin: safest approach to preserve food products. *Indian J Microbiol.* 2009;49;3:204-211.
 7. Riley MA, Wertz JE. Bacteriocin diversity: ecological and evolutionary perspectives. *Biochimie.* 2002;84;5-6:357-364.
 8. Frederico P. Colicins. *Annu Rev Microbiol.* 1957;11;1:7-22.
 9. Cleveland J, Montville TJ, Nes IF, Chikindas ML. Bacteriocins: safe, natural antimicrobials for food preservation. *Int J Food Microbiol.* 2001;71;1:1-20.
 10. Shin JM, Gwak JW, Kamarajan P, Fenno JC, Rickard AH, Kapila YL. Biomedical applications of nisin. *J Appl Microbiol.* 2016;120;6:1449-1465.
 11. Cotter PD, Ross RP, Hill C. Bacteriocins-a viable alternative to antibiotics? *Nat Rev Microbiol.* 2013;11;2:95-105.
 12. Kaur S, Kaur S. Bacteriocins as potential anticancer agents. *Front Pharmacol.* 2015 ;6; 272.
 13. Yang SC, Lin CH, Sung CT, Fang JY. Antibacterial activities of bacteriocins: Application in foods and pharmaceuticals. *Front Microbiol.* 2014;5;241.
 14. Stein T. Whole-cell matrix-assisted laser desorption/ionization mass spectrometry for rapid identification of bacteriocin/lantibiotic-producing bacteria. *Rapid Commun Mass Spectrom.* 2008;22;8:1146-1152.
 15. Wosinska L, Cotter PD, O'Sullivan O, Guinane C. The potential impact of probiotics on the gut microbiome of athletes. *Nutrients.* 2019;11;10.
 16. Ibrahim OO. Classification of antimicrobial peptides bacteriocins, and the nature of some bacteriocins with potential applications in food safety and bio-pharmaceuticals. *EC Microbiol.* 2019;15:591-608.
 17. Moll GN, Konings WN, Driessen AJM. Bacteriocins: mechanism of membrane insertion and pore formation. *Lact acid Bact Genet Metab Appl.* 1999;76;1-4:185-198.
 18. Nissen-Meyer J, Oppegård C, Rogne P, Haugen HS, Kristiansen PE. Structure and mode-of-action of the two-peptide (class-IIb) bacteriocins. *Probiotics Antimicrob Proteins.* 2010;2;1:52-60.
 19. Diep DB, Skaugen M, Salehian Z, Holo H, Nes IF. Common mechanisms of target cell recognition and immunity for class II bacteriocins. *Proc Natl Acad Sci.* 2007;104;7:2384-2389.
 20. Lee H-J, Kim H-Y. Lantibiotics, class I bacteriocins from the genus *Bacillus*. *J Microbiol Biotechnol.* 2011;21;3:229-235.
 21. Blond A, Cheminant M, Destoumieux-Garzón D, et al. Thermolysin-linearized microcin J25 retains the structured core of the native macrocyclic peptide and displays antimicrobial activity. *Eur J Biochem.* 2002;269;24:6212-6222.
 22. Baquero F, Moreno F. The microcins. *FEMS Microbiol Lett.* 1984;23;2-3:117-124.
 23. Pons AM, Lanneluc I, Cottenceau G, Sable S. New developments in non-post translationally modified microcins. *Biochimie.* 2002;84;5-6:531-537.
 24. Lagos R, Tello M, Mercado G, Garcia V, Monasterio O. Antibacterial and Antitumorigenic Properties of Microcin E492, a Pore- Forming Bacteriocin. *Curr Pharm Biotechnol.* 2009;10;1:74-85.
 25. Mathavan I, Beis K. The role of bacterial membrane proteins in the internalization of microcin MccJ25 and MccB17. In: *Biochemical Society Transactions.* 2012;40:1539-1543.
 26. Riley MA. Bacteriocins, biology, ecology, and evolution. *Encycl Microbiol.* 2009:32-44.
 27. Davies JK, Reeves P. Genetics of resistance to colicins in *Escherichia coli* K-12: cross-resistance among colicins of group A. *J Bacteriol.* 1975;123;1:102-117.
 28. Davies JK, Reeves P. Genetics of resistance to colicins in *Escherichia coli* K-12: cross-resistance among colicins of group B. *J Bacteriol.* 1975;123;1:96-101.
 29. Klaenhammer TR. Genetics of bacteriocins produced by lactic acid bacteria. *FEMS Microbiol Rev.* 1993;12;1-3:39-85. doi:10.1111/j.1574-6976.1993.tb00012.x
 30. Cotter PD, Hill C, Ross RP. Food microbiology: Bacteriocins: Developing innate immunity for food. *Nat Rev Microbiol.* 2005;3;10:777-788.
 31. Arnison PG, Bibb MJ, Bierbaum G, et al. Ribosomally synthesized and post-translationally modified peptide natural products: Overview and recommendations for a universal nomenclature. *Nat Prod Rep.* 2013;30;1:108-160.
 32. Soltani S, Hammami R, Cotter PD, et al. Bacteriocins as a new generation of antimicrobials: Toxicity aspects and regulations. *FEMS Microbiol Rev.* 2021;45;1.
 33. Wachsman MÓB, Farías ME, Takeda E, et al. Antiviral activity of enterocin CRL35 against herpesviruses. *Int J Antimicrob Agents.* 1999;12;4:293-299.
 34. Wachsman MB, Castilla V, De Ruiz Holgado AP, De Torres RA, Sesma F, Coto CE. Enterocin CRL35 inhibits late stages of HSV-1 and HSV-2 replication in vitro. *Antiviral Res.* 2003;58;1:17-24.

35. Todorov SD, Wachsman MB, Knoetze H, Meincken M, Dicks LMT. An antibacterial and antiviral peptide produced by *Enterococcus mundtii* ST4V isolated from soya beans. *Int J Antimicrob Agents*. 2005;25;6:508-513.
36. Cavicchioli VQ, Carvalho OV de, Paiva JC de, Todorov SD, Silva Júnior A, Nero LA. Inhibition of herpes simplex virus 1 (HSV-1) and poliovirus (PV-1) by bacteriocins from *Lactococcus lactis* subsp. *lactis* and *Enterococcus durans* strains isolated from goat milk. *Int J Antimicrob Agents*. 2018;51;1:33-37.
37. Todorov SD, Wachsman M, Tomé E, et al. Characterisation of an antiviral pediocin-like bacteriocin produced by *Enterococcus faecium*. *Food Microbiol*. 2010;27;7:869-879.
38. Ermolenko EI, Desheva YA, Kolobov AA, Kotyleva MP, Sychev IA, Suvorov AN. Anti-Influenza Activity of Enterocin B In vitro and Protective Effect of Bacteriocinogenic Enterococcal Probiotic Strain on Influenza Infection in Mouse Model. *Probiotics Antimicrob Proteins*. 2019;11;2:705-712.
39. Férrir G, Petrova MI, Andrei G, et al. The Lantibiotic Peptide Labyrinthopeptin A1 Demonstrates Broad Anti-HIV and Anti-HSV Activity with Potential for Microbicidal Applications. *PLoS One*. 2013;8;5.
40. Torres NI, Noll KS, Xu S, et al. Safety, Formulation and In Vitro Antiviral Activity of the Antimicrobial Peptide Subtilosin Against Herpes Simplex Virus Type 1. *Probiotics Antimicrob Proteins*. 2013;5;1:26-35.
41. Quintana VM, Torres NI, Wachsman MB, Sinko PJ, Castilla V, Chikindas M. Antiherpes simplex virus type 2 activity of the antimicrobial peptide subtilosin. *J Appl Microbiol*. 2014;117;5:1253-1259.
42. Tabata T, Pettitt M, Puerta-Guardo H, et al. Zika Virus Targets Different Primary Human Placental Cells, Suggesting Two Routes for Vertical Transmission. *Cell Host Microbe*. 2016;20;2:155-166.
43. Richard AS, Zhang A, Park SJ, Farzan M, Zong M, Choe H. Virion-associated phosphatidylethanolamine promotes TIM1-mediated infection by Ebola, dengue, and West Nile viruses. *Proc Natl Acad Sci U S A*. 2015;112;47:14682-14687.
44. Lee M, Yang J, Park S, et al. Micrococcin P1, a naturally occurring macrocyclic peptide inhibiting hepatitis C virus entry in a pan-genotypic manner. *Antiviral Res*. 2016;132:287-295.
45. Serkedjieva J, Danova S, Ivanova I. Antiinfluenza virus activity of a bacteriocin produced by *Lactobacillus delbrueckii*. In: *Applied Biochemistry and Biotechnology - Part A Enzyme Engineering and Biotechnology* Springer. 2000;88:285-298.