

ORIGINAL ARTICLE

Antibiotic Resistance Pattern of *Propionibacterium acnes* Isolated from Patients with Acne Vulgaris at the Dermatology Clinics of Kasr Al-Ainy Teaching Hospital Egypt

¹Rasha A. Elfekki, ²Nesrin Samir, ³Nadia Hafez Ouda, ³Asmaa S. Hegab*

¹Department of Medical Microbiology and Immunology, Helwan General Hospital, Cairo11735, Egypt

²Department of Dermatology, Faculty of Medicine, Cairo University, Cairo11562, Egypt

³Department of Medical Microbiology and Immunology, Faculty of Medicine, Cairo University, Cairo11562, Egypt

ABSTRACT

Key words:

P. acnes
(*Propionibacterium acnes*),
Acne Vulgaris, antibiotic
resistance, clindamycin,
erythromycin,
trimethoprim-
sulfamethoxazole

*Corresponding Author:

Asmaa Hegab
Department of Medical
Microbiology and
Immunology, Cairo University,
Egypt
Tel.: 01005195939
asmahgab@kasralainy.edu.eg

Background: *Propionibacterium acnes* (*P. acnes*) has been implicated in the pathogenesis of acne vulgaris since the beginning of the last century. Over several decades, topical and systemic antibiotics have been the main line of treatment for acne vulgaris. However, in the present era of increased antibiotic usage, resistant strains have emerged. **Objectives:** The aim of this study is to determine the antibiotic resistance pattern of *P. acnes* isolated from patients with acne vulgaris at the dermatology clinics of Kasr Al-Ainy teaching Hospital. **Methodology:** Specimens from 100 patients with acne vulgaris were extracted from the pustules, taken by sterile cotton swabs and transported by thioglycolate media. Each swab was inoculated onto two blood agar plates, one incubated aerobically at 37°C for 24h and the other anaerobically for one week. *P. acnes* was identified by Gram stain and biochemical tests. Their sensitivity to doxycycline, erythromycin, clindamycin, tetracycline, azithromycin and trimethoprim-sulfamethoxazole was determined on Muller Hinton media by disc diffusion method. Results were interpreted according to the National Committee for Clinical Laboratory Standards (NCCLS) guidelines. **Results:** A total of 44 *P. acnes* isolates were identified from 100 patients with acne vulgaris, out of which 22.7% were resistant to clindamycin, 11.4% were resistant to trimethoprim-sulfamethoxazole and 9% were resistant to erythromycin. Resistance to doxycycline, tetracycline or azithromycin was not detected. Trimethoprim-sulfamethoxazole showed statistically significant difference in the resistance pattern compared to patient's sex ($p = 0.029$) and to receiving previous treatment ($p = 0.018$). **Conclusion:** *P. acnes* is prevalent in patients with acne vulgaris (44%) and resistant strains are detected especially in those who have received previous therapy for more than 2 weeks (68%). It is recommended that dermatologists and family physicians follow the guidelines for proper management of acne, with the judicious use of antibiotics, in order to prevent antibiotic resistance.

INTRODUCTION

Acne vulgaris is a chronic inflammatory disorder of pilosebaceous follicles affecting more than 85% of adolescents and young adults¹. It is characterized by a variety of eruptions including comedones, erythematous papules, pustules, and sometimes nodules, frequently followed by scarring². Acne is not an infectious disease, but

P. acnes, *Staphylococcus epidermidis* (*S. epidermidis*), and *Staphylococcus aureus* (*S. aureus*), organisms residing on the surface of skin and pilo sebaceous ducts may trigger infection. Pathogenesis of acne may be attributed to inflammation, hyperkeratinisation of the sebaceous duct, high sensitivity to circulating androgens, and bacterial colonization². Widespread and long-term use of

antibiotics such as erythromycin, clindamycin, and tetracycline has led to the development of *P. acnes* resistance³. The increasing prevalence of antimicrobial resistance in *P. acnes* poses a significant challenge to successful treatment outcomes in acne patients⁴.

This study was undertaken to examine the percentage of *P. acnes* within the bacteriological profile isolated from acne vulgaris patients and to ascertain isolated *P. acnes* antibiotic resistance pattern.

METHODOLOGY

A cross-sectional study was undertaken in the Outpatient Clinic of Dermatology Department, Kasr El-Ainy Teaching Hospital, Cairo University; over a period of two years between 2013 and 2014. Inclusion criteria were: (1) 14-25 years of age; and (2) mild-to-severe

acne vulgaris. Exclusion criteria were: (1) oral or topical antibiotic in the past two weeks; (2) other facial skin diseases. A total of 100 patients with pustular acne vulgaris were included after informed consent was obtained. The study was approved by the ethical Committee of Kasr El-Ainy Teaching Hospital, Cairo University. Detailed history and clinical examination were carried out.

Skin sampling:

Samples were collected from pustular acne lesions of 100 patients. The skin was first wiped out with 70% ethanol and the samples were taken using sterile disposable cotton swabs after rupture of the pustule with sterile hypodermic needle. The swabs were inserted into anaerobic transport medium [thioglycolate broth] (Merck, Darmstadt, Germany)⁵. Samples were transported immediately to Medical Microbiology and Immunology Department, Cairo University, to be processed within 24 hours of collection. All specimens were subjected to aerobic and anaerobic culture.

Bacteriological study:

The specimens were inoculated onto blood agar supplemented with hemin and menadione (Thermo Fisher Scientific, UK). Plates were incubated at 37°C under aerobic condition for 24 hrs and under anaerobic condition (AnaeroGen; Oxoid Ltd, Basingstoke, UK) for one week; and examined for growth. After one week if culture was negative for *P. acnes*, a subculture was done from the thioglycolate medium and incubated for another week anaerobically⁵.

Anaerobic culture was performed using the Gaspak system, Oxoid AnaeroGen (OXOID Ltd, Wade road, England). The AnaeroGen sachet will absorb the atmospheric oxygen thus reducing the oxygen level in the jar to below 1% within 30 minutes and results in 9-

13% carbon dioxide simultaneously. Unlike other gas generating systems, AnaeroGen does not produce hydrogen gas and therefore, does not require a catalyst. Furthermore, no addition of water is needed to activate the reaction⁶.

Oxoid anaerobic indicator was used to ensure anaerobiosis. It is a disposable sachet containing a strip of inert material saturated with a resazurin solution. In use, a color change from pink to white indicates redox condition suitable for the growth of most anaerobic organisms. If after incubation the indicator strip has not changed color, the anaerobic culture must be repeated⁵.

Identification:

Aerobic bacteria were identified by colony morphology, Gram stain and standard biochemical tests⁷. *P. acnes* strains were identified as small, white to pink, shiny to opaque colonies with entire margins, grown anaerobically, showing Gram positive, non-spore forming, slender, slightly curved rods with positive indole, catalase, and glucose fermentation tests as well as negative bile esculine and urease tests⁸.

Antibiotic susceptibility:

Antibiotic susceptibility of *P. acnes* isolates was performed by disc diffusion method using commercially available discs (antibiotic discs; Oxoid, Basingstoke, UK) according to the NCCLS guidelines. Standardized Kirby-Bauer disc diffusion method was performed on Mueller-Hinton agar (LAB™, Lancashire, England). Inocula were adjusted to a 0.5 MacFarland standard from an anaerobic pure culture on blood agar. Mueller-Hinton agar plates were inoculated and incubated at 37°C for 5-7 days anaerobically. Results were interpreted according to the NCCLS guidelines (table 1)⁹.

Table 1: Antibiotic discs used to test for *P. acnes* susceptibility⁹

	Concentration	Susceptible zone diameter (mm)		
		Resistant	Intermediate	Sensitive
Clindamycin	2 µg	14	15 - 20	21
Trimethoprim-Sulfamethoxazole	25 µg	10	11 - 15	16
Erythromycin	15 µg	13	14 - 22	23
Tetracycline	30 µg	14	15 - 18	19
Doxycycline	30 µg	12	13 - 15	16
Azithromycin	15 µg	13	14 - 17	18

Statistical Analysis:

The statistical package SPSS version 15 was used to code and enter the data. The data was summarized using descriptive statistics: mean, standard deviation, minimal and maximum values for quantitative variables as well as number and percentage for qualitative values. Statistical differences between groups were tested using Chi Square test for qualitative variables and independent sample t test for quantitative normally

distributed variables. *P* values ≤ 0.05 were considered statistically significant.

RESULTS

The demographic and clinical data of 100 acne patients are shown in table 2. All the patients were between 14 and 25 years of age, with a mean age of 19.6 ± 2.9 years. The majority of patients were between

20 and more years of age (57%), with a predominance of females (72%). The disease duration exceeded 2 years in 57% of patients. Thirty-one percent of patients had history of previous anti-acne treatment. There was no significant difference in the detection of *P. acnes* in relation to the patients' age, sex or duration of the disease. However, there was a statistically significant difference in patients taking previous treatment. *P. acnes* was more prevalent in patients who received previous medical treatment for more than 2 weeks ($P = 0.001$) (table 2).

Table 2: General characteristics of patients with *P. acnes*

	No.	%	P value
Age (years)			
< 20	20 / 43	46.5%	0.660
≥ 20	24 / 57	42.1%	
Sex			
Female	34 / 72	47.2%	0.298
Male	10 / 28	35.7%	
Duration of disease (years)			
< 2	21 / 43	48.8%	0.397
> 2	23 / 57	40.4%	
Previous treatment			
Yes	21 / 31	67.7%	0.001
No	23 / 69	33.3%	

The bacteriological profiles of the acne patients are shown in Table 3. Of 100 samples processed, 91% samples showed aerobic growth and 47% showed anaerobic growth. *S. epidermidis* (61; 61%) was the predominant aerobe, followed by *S. aureus* (28; 28%) and *Candida* (2; 2%). Among the anaerobes, *Propionibacterium* species were isolated from 47 samples, with *P. acnes* as the predominant species (44;

44%); *Propionibacterium granulosum* (indole test: negative) was detected in 3% of samples (table 3). Out of the 44 *P. acnes* isolates, only one was isolated in pure form and 43 were isolated in mixed form (table 4).

Table 3: Isolated microorganisms from the studied patients

	%
Aerobic	
<i>S. aureus</i>	28
<i>S. epidermidis</i>	61
<i>Candida</i>	2
Anaerobic	
<i>P. acnes</i>	44
<i>P. granulosum</i>	3

Table 4: Percentage of *P. acnes* isolated from patients suffering from acne vulgaris

	%
Mixed growth with <i>P. acnes</i>	43
Pure <i>P. acnes</i>	1
No growth	5
Bacteria other than <i>P. acnes</i>	51

Antibiotic susceptibility pattern:

10 *P. acnes* strains showed resistance to clindamycin (22.7%), 5 strains were resistant to trimethoprim-sulfamethoxazole (11.4%) and 4 strains were resistant to erythromycin (9.1%). Three isolates showed resistance to both clindamycin and erythromycin. No resistance was observed to azithromycin (0%), doxycycline (0%) nor tetracycline (0%) (table 5).

Table 5. Antibiotic susceptibility pattern of isolated *P. acnes*

Antibiotic	<i>P. acnes</i> susceptibility pattern (no. = 44)		
	Susceptible No. (%)	Intermediate No. (%)	Resistant No. (%)
Clindamycin	31 (70.5%)	3 (6.8%)	10 (22.7%)
Trimethoprim- Sulfamethoxazole	35 (79.5%)	4 (9.1%)	5 (11.4%)
Erythromycin	37 (84.1%)	3 (6.8%)	4 (9.1%)
Tetracycline	43 (97.7%)	1 (2.3%)	0 (0%)
Doxycycline	44 (100%)	0 (0%)	0 (0%)
Azithromycin	44 (100%)	0 (0%)	0 (0%)

There were no statistically significant differences in the antibiotic resistance pattern of isolated *P. acne* in relation to patients' age, sex, disease duration and previous treatment. However, there was a statistically

significant difference in the resistance pattern of trimethoprim-sulfamethoxazole when compared to sex of patients ($P = 0.029$) and history of taking previous treatment ($P = 0.018$) (table 6) (Figures 1, 2).

Table 6: Prevalence of antibiotic resistance to *P. acnes* according to age, sex, disease duration and previous treatment

	Clindamycin	Trimethoprim – Sulfamethoxazole	Erythromycin	Tetracycline	Doxycycline	Azithromycin
Age (years)						
< 20	4 / 20 (20%)	1 / 20 (5%)	2 / 20 (10%)	0 / 20(0%)	0 / 20(0%)	0/20(0%)
≥ 20	6 / 24 (25%)	4 / 24(16.7%)	2 / 24(8.3%)	0 / 24(0%)	0 / 24(0%)	0 / 24(0%)
<i>P</i> value	0.817	0.478	0.261	0.000	0.000	0.000
Sex						
Female	10 / 34 (29.4%)	2 / 34(5.9%)	3/34(8.8%)	0/34(0%)	0/34(0%)	0 / 34(0%)
Male	0 / 10 (0%)	3 /10(30%)	1 / 10(10%)	0 / 10(0%)	0 / 10(0%)	0 / 10(0%)
<i>P</i> value	0.066	0.029	0.623	0.000	0.000	0.000
Duration of disease (years)						
< 2	4 / 21 (19%)	3/21(14.3%)	2 / 21(9.5%)	0 / 21(0%)	0 / 21(0%)	0 / 21(0%)
> 2	6 / 23(26.1%)	2/ 23(8.7%)	2 / 23(8.7%)	0 / 23(0%)	0 / 23(0%)	0 / 23(0%)
<i>P</i> value	0.713	0.832	0.230	0.000	0.000	0.000
Previous treatment						
Yes	6 / 21(28.6%)	4 / 21(19%)	4 / 21(19%)	0 / 21(0%)	0 / 21(0%)	0 / 21(0%)
No	4 / 23(17.4%)	1 / 23(4.3%)	0 / 23(0%)	0 / 23(0%)	0 / 23(0%)	0 / 23(0%)
<i>P</i> value	0.187	0.018	0.061	0.000	0.000	0.000

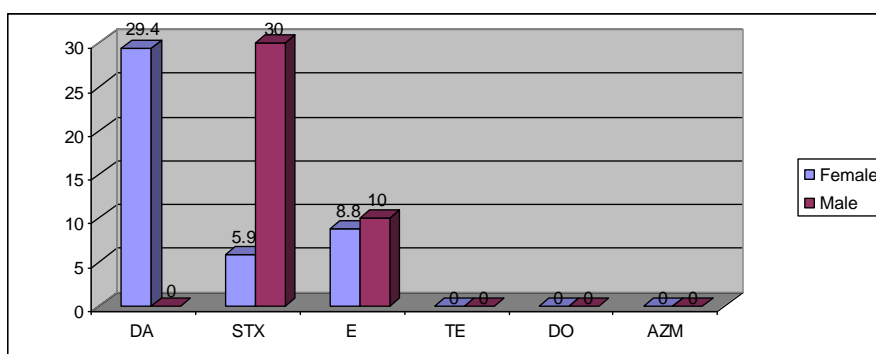


Fig. 1: Antibiotic resistance pattern in relation to sex

DA: Clindamycin; SXT: Trimethoprim-sulfamethoxazole; E; Erythromycin; TE: Tetracycline; DO: Doxycycline, AZM: Azithromycin.

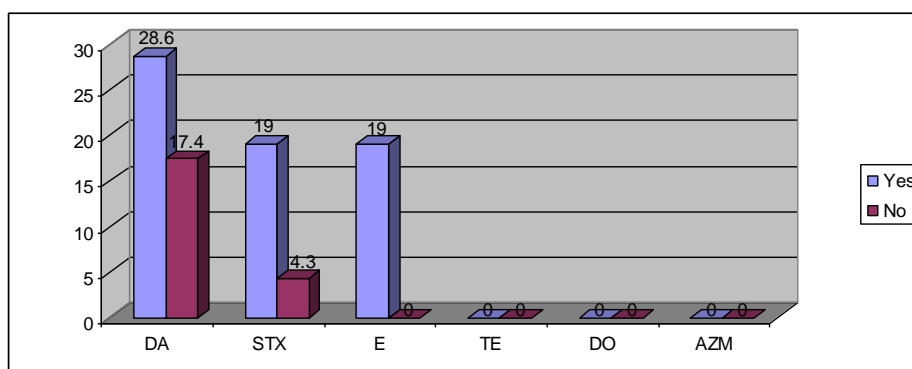


Fig. 2: Antibiotic resistance pattern in relation to previous treatment

DA: Clindamycin; SXT: Trimethoprim-sulfamethoxazole; E; Erythromycin; TE: Tetracycline; DO: Doxycycline, AZM: Azithromycin.

DISCUSSION

Cutaneous *P. acnes* has been implicated in acne, although their role in inflammation is still poorly understood. There is widespread resistance in *P. acnes* due to over use of topical and systemic antibiotics for treatment of acne vulgaris¹⁰. The difference in the *in vitro* antibiotic susceptibility patterns of *P. acnes* among different countries is not clear; however it may be attributed to the different antibiotic prescribing habits and/or different ethnicity of the patients as well as genetic mutations¹¹. Different studies have used various interpretative criteria to estimate the resistance among *P. acnes* isolates to different anti-acne drugs. The available data on antibiotic resistance among these strains are deficient; also, no standard criterion is available for determining the resistance among anti-acne drugs in *P. acnes*. Virtually, drug susceptibility is not prescribed, owing to the slow growth of the bacteria and the cost and complexity of testing methods. However, it is important to get resistance information so that correct therapeutic decisions can be made, particularly in resistant cases not responding to routine therapy².

Our study showed a high prevalence of acne in female patients (72%), which was in agreement with the studies done by Hassan *et al.* (73%)¹², Moon *et al.* (68%)¹³, Zandi *et al.* (64%)⁵ and Zhu T *et al.* (59%)¹⁴. Forty three percent of our patients had a disease duration of less than 2 years while 57% had a disease course of more than 2 years. In agreement, Moon *et al.*¹³ reported 38% and 60% respectively and Zhu T *et al.*¹⁴ reported 20% and 80% respectively. In contrast, Sardana *et al.*¹⁵ reported 52% and 46% respectively. In the present study, 69% of patients did not receive any anti-acne drugs. Similarly, this was reported by Moon *et al.* (73%)¹³, Zandi *et al.* (65%)⁵ and Zhu T *et al.* (64%)¹⁵. In contrast, Mendoza *et al.*⁴ reported that 75% of patients received previous anti-acne treatment.

Out of the 100 patients diagnosed with acne vulgaris, *P. acnes* represented 44% of isolates when cultured anaerobically. In agreement with our results, *P. acnes* was detected in 41%, 35%, 35% and 32% of the samples of the following studies Zandi *et al.*⁵, Hassanzadeh *et al.*¹⁶, Hassan *et al.*¹² as well as Dhillon and Varshney¹⁷ respectively. Other studies reported a higher percentage of isolated *P. acnes* as Schafer *et al.*¹⁸ (96%) as well as Abdel Fattah and Darwish¹⁹ (88%). This may be due to the greater number of patients included in their study as compared to our study. In contrast, Moon *et al.*¹³ reported a lower percentage (20%) which may be attributed to the short duration of the study.

In this study, out of the 44 *P. acnes* isolates, only one was isolated in pure form (mixed growth was detected in 43%). *S. epidermidis* (61%) and *S. aureus* (28%) were the most prevalent bacteria detected

aerobically. In agreement with this results, Hassanzadeh *et al.*¹⁶ reported that *S. epidermidis* were detected in 53% of isolates. Moreover, Nishijima *et al.*²⁰ reported that *S. epidermidis* was detected in more than 50% of their isolates and Hassan *et al.*¹² reported the detection of *S. epidermidis* in 44% of the isolates. In contrast, other studies reported that the most frequent bacterium isolated from patients with acne was *S. aureus*. Dhillon and Varshney¹⁷ detected *S. aureus* in 41% and *S. epidermidis* in 20%. Toyoda and Morohashi²¹ detected *S. aureus* in 48% and *S. epidermidis* in 23%. This difference in microbial profile in our study could be explained by the variations in geographical location, host factors, and antibiotic usage.

Moreover, we studied the significance of *P. acnes* detection in relation to the patients' age, sex, disease duration and history of previous treatment. The only statistically significant relation we found was the increased incidence of *P. acnes* (68%) in patients receiving previous treatment for more than 2 weeks ($p=0.001$). Similarly, Tan *et al.*²² and Schafer *et al.*¹⁸ reported a prevalence of 75% and 70% respectively. Thus, confirming that *P. acnes* is one of the most common causes of resistant acnes lesions¹⁸. However, other studies as Abdel Fattah and Darwish¹⁹ reported a lower frequency (33%).

Regarding the susceptibility pattern, we found that 22.7% of the isolated *P. acnes* were resistant to clindamycin, 11.4% were resistant to trimethoprim-sulfamethoxazole and 9% were resistant to erythromycin. Several studies in different countries reported clindamycin resistance: In India, 90% by Sardana *et al.*¹⁵, in Egypt, 85% by Hassan *et al.*¹² and 65% by Abdel Fattah and Darwish¹⁹, in Hong Kong, 53.5% by Luk *et al.*¹¹, in Iran, 50% by Zandi *et al.*⁵, in Mexico, 35% by Gonzalez *et al.*²³, in Korea, 26.7% by Moon *et al.*¹³ and in Japan, 10.3% by Ishida *et al.*²⁴. Our percentage of clindamycin resistance was lower than most studies, probably due to the lower number of isolated *P. acnes* (44%). In addition, different geographical regions may affect the distribution of bacteria involved in acne vulgaris¹². However, in agreement with all these studies, we found that clindamycin resistance was higher than erythromycin resistance within the isolated *P. acnes*. This is possible due to the fact that clindamycin is the most commonly prescribed antibiotic for patients with acne, in most communities including ours¹².

In addition, 3 out of the 44 *P. acnes* isolates (7%) had cross resistance to both clindamycin and erythromycin. Another Egyptian study also reported cross-resistance to clindamycin and erythromycin in 20 out of 98 patients (20%)¹⁹. Moreover, a study from Greece similarly reported cross-resistance between the two antibiotics in 23 out of 79 patients (29%)²⁵. The cross resistance to clindamycin and erythromycin found

in our study could be attributed to the use of both antibiotics as topical and systemic solutions for acne patients¹⁹. The underlying mechanism of erythromycin and clindamycin cross-resistance was elucidated by Ross *et al.* who identified four phenotypes with cross susceptibility to macrolide, lincosamide and streptogramin B (MLS) antibiotics. Genetic mutations occur mainly in 23S rRNA, and strains that possess the *erm* (X) resistance gene are highly resistant to MLS antibiotics¹⁰.

In this study, 11.4% of isolated *P. acnes* were resistant to trimethoprim-sulfamethoxazole. This may be attributed to its high usage in sinusitis. Similarly, the following studies reported resistance to trimethoprim-sulfamethoxazole: 26.3% by Schafer *et al.*¹⁸, 15% by Ross *et al.*¹⁰ and 8% by Oprica *et al.*²⁶. On the other hand, Gonzalez *et al.* reported 68% trimethoprim-sulfamethoxazole resistance²³. This higher percentage could be attributed to the overuse of this antibiotic by physicians, as well as the fact that it is usually sold in Mexico and other countries as an over-the-counter medication for other infections²³.

Regarding tetracycline and doxycycline susceptibility, all *P. acnes* isolates in this study were highly susceptible to both antibiotics. Our results are also consistent with those of Giannopoulos *et al.* who reported no resistance to tetracycline in Greece²⁵, Schafer *et al.* in Chile¹⁸, Ross *et al.* in Italy and Hungary¹⁰ and Doğan *et al.* in Turkey²⁷. This is possibly attributed to the fact that topical cyclines are not usually prescribed for patients with *acne vulgaris* which in turn decreases the emergence of *P. acnes* cycline-resistant strains¹⁹. The European guidelines for acne recommend tetracycline as a first line treatment in acne patients²⁸. Tetracycline and doxycycline resistance patterns were detected worldwide. In Egypt (18% & 6% respectively)¹⁹, in Hong Kong (16.3% for both)¹¹, in Mexico (14% & 20% respectively)²³, in Singapore (11.5% for both)²¹ in France (9.5% for both)²⁹, in Japan (2% for both)²³ and in southwest China (3% & 1% respectively)¹⁴. In contrast, a higher tetracycline resistance pattern was detected in Iran (35%)⁵ and in India (30%)¹⁵, due to its high usage in these countries.

In the present study, all isolates were susceptible to azithromycin. The Egyptian study by Abdel Fattah and Darwish reported a low percentage of azithromycin resistance (5%) and suggested azithromycin as an important line for acne treatment in Egyptian patients¹⁸. Similarly, no azithromycin resistance was reported in Iran⁵. However, in contrast to our findings, higher resistance to azithromycin was reported: In China 59% by Zhu T *et al.*¹⁴, in Mexico 82% by Gonzalez *et al.*²³, and in India 100% by Sardana *et al.* suggesting it is probably attributed to a cross resistance mechanism similar to that existing between clindamycin and erythromycin, and recommended that further studies are required to exclude the possibility of cross-resistance

between this macrolide antibiotic and lincosamides (clindamycin)¹⁵.

When trying to correlate the prevalence of resistant *P. acnes* isolates with age, disease duration and previous treatment, no statistically significant results were found. A significant difference was found when correlating previous acne treatment with trimethoprim-sulfamethoxazole resistance ($p=0.018$). In agreement, Schafer *et al.*¹⁸, Abdel Fattah and Darwish¹⁹, Nord and Oprica³⁰ and Ross *et al.*¹⁰ reported that resistant *P. acnes* were more prevalent in patients who received previous acne treatment. No resistant *P. acnes* isolates were detected in patients who did not receive previous treatment.

Owing to the fact that acne often requires long-term treatment, there is a concern that *P. acnes* resistance may be associated with the development of resistance in other organisms such as *S. aureus* and *S. epidermidis*. Thus, it is important that antibiotics be used judiciously, particularly in such a common condition as *acne vulgaris*¹³.

CONCLUSION

We concluded that highest resistance rates were detected with clindamycin, followed by trimethoprim-sulfamethoxazole and erythromycin (22.7%, 11.4% and 9% respectively), while all *P. acnes* isolates were susceptible to azithromycin and doxycycline and almost all of them (98%) were susceptible to tetracycline. This can change the strategy for treatment of acne.

Acknowledgements

We acknowledge the Medical microbiology and immunology department, Faculty of Medicine, Cairo University, Egypt, for their support.

Conflicts of interest: The authors declare that they have no financial or non-financial conflicts of interest related to the work done in the manuscript.

- Each author listed in the manuscript had seen and approved the submission of this version of the manuscript and takes full responsibility for it.
- This article had not been published anywhere and is not currently under consideration by another journal or a publisher.

REFERENCES

1. Hanna S, Sharma J, Klotz J. Acne vulgaris: More than skin deep. *Dermatol Online J* 2003; 9: 8.
2. Biswal I, Gaiind R, Kumar N, Mohanty S, Manchanda V, Khunger N, et al. *In vitro* antimicrobial susceptibility patterns of *Propionibacterium acnes* isolated from patients

- with acne vulgaris. *J Infect Dev Ctries* 2016; 10:1140-1145.
3. Leyden J, Del Rosso J, Webster G. Clinical considerations in the treatment of acne vulgaris and other inflammatory skin disorders: Focus on antibiotic resistance. *Cutis* 2007; 79: 9-25.
 4. Mendoza N, Hernandez P, Tying S. Antimicrobial susceptibility of *Propionibacterium acnes* isolates from acne patients in Colombia. *Int J Dermatol* 2013; 52: 688-692.
 5. Zandi S, Vares B, Abdollahi H. Determination of microbial agents of acne vulgaris and *Propionibacterium acnes* antibiotic resistance in patients referred to dermatology clinics in Kerman, Iran. *Jundishapur J Microbiol* 2011; 4: 17-22.
 6. Shahin M, Jamal W, Verghese T. Comparative evaluation of anoxomat and conventional anaerobic GasPak jar systems for the isolation of anaerobic bacteria. *Med Princ Pract* 2003; 12: 81-86.
 7. Normanno G, Firinu A, Virgilio S. Coagulase-positive staphylococci and *Staphylococcus aureus* in food products marketed in Italy. *Int J Food Microbiol* 2005; 98: 73-79.
 8. Cauich-Sanchez P, Alatraste-Mondragon F, Garcia-Cano E. Identification of anaerobic non spore-forming Gram-positive bacilli by biochemical tests and gas-liquid chromatography. *Revista latinoamericana de microbiologia* 2001; 43: 27-35.
 9. National Committee for Clinical Laboratory Standards. Performance standards for antimicrobial disk susceptibility tests. Approved standard, 8th ed 2003; 23(1).
 10. Ross JI, Snelling AM, Eady EA, Cove JH, Cunliffe WJ, Leyden JJ, et al. Phenotypic and genotypic characterization of antibiotic-resistant *Propionibacterium acnes* isolated from acne patients attending dermatology clinics in Europe, the USA, Japan and Australia. *Br J Dermatol* 2001; 144: 339-346.
 11. Luk NM, Hui M, Lee HC. Antibiotic-resistant *Propionibacterium acnes* among acne patients in a regional skin centre in Hong Kong. *J Eur Acad Dermatol Venereol* 2013; 27: 31-36.
 12. Hassan IA, Hassan MA, Embarek MS, Attallah DA, ElMokhtar MA, Alaa Eldin GM. Antibiotic susceptibility patterns of *Propionibacterium acnes* isolated from acne vulgaris in Assiut University hospitals, Egypt. *Egypt J Med Microbiol* 2015; 24(4): 67-72.
 13. Moon S, ROH H, Kim Y. Antibiotic resistance of microbial strains isolated from Korean acne patients. *Journal of Dermatology* 2012; 39: 1-5.
 14. Zhu T, Zhu W, Wang Q, He L, Wu W, Liu J, et al. Antibiotic susceptibility of *Propionibacterium acnes* isolated from patients with acne in a public hospital in Southwest China: prospective cross-sectional study. *BMJ* (2019). <http://bmjopen.bmj.com/>
 15. Sardana K, Gupta T, Kumar B, Gautam HK, Garg VK. A cross-sectional pilot study of antibiotic resistance in *Propionibacterium acnes* strains in Indian acne patients using 16S RNA polymerase chain reaction: A comparison among treatment modalities including antibiotics, benzoyl peroxide and isotretinoin. *Indian J Dermatol* 2016; 61(1): 45-52.
 16. Hassanzadeh P, Bahmani M, Mehrabani D. Bacterial resistance to antibiotics in acne vulgaris: An *in vitro* study. *Indian J Dermatol* 2008; 53(3): 122-124.
 17. Dhillon K, Varshney KR. Study of microbiological spectrum in acne vulgaris: An *in vitro* study. *Sch J App Med Sci* 2013; 1: 724-727.
 18. Schafer F, Fich F, Lam M. Antimicrobial susceptibility and genetic characteristics of *Propionibacterium acnes* isolated from patients with acne. *Int J Dermatol* 2013; 52: 418-425.
 19. Abdel Fattah NS, Darwish YW. *In vitro* antibiotic susceptibility patterns of *Propionibacterium acnes* isolated from acne patients: An Egyptian university hospital-based study. *J Eur Acad Dermatol Venereol* 2013; 27: 1546-1551.
 20. Nishijima S, Kurokawa I, Katoh N. The bacteriology of acne vulgaris and antimicrobial susceptibility of *Propionibacterium acnes* and *Staphylococcal epidermidis* isolated from acne lesions. *Int J Dermatol* 2000; 27: 318-323.
 21. Toyoda M, Morohashi M. An overview of topical antibiotics for acne treatment. *Dermatology*; 196: 130-134.
 22. Tan HH, Tan AW, Barkham T. Community-based study of acne vulgaris in adolescents in Singapore. *Br J Dermatol* 2007; 157: 547-551.
 23. Gonzalez R, Welsh O, Ocampo J (2010) *In vitro* antimicrobial susceptibility of *Propionibacterium acnes* isolated from acne patients in Northern Mexico. *Int J Dermatol*; 49: 1003-1007.
 24. Ishida N, Nakaminami H, Noguchi N. Antimicrobial susceptibilities of *Propionibacterium acnes* isolated from patients with acne vulgaris. *Microbiol Immunol* 2008; 52: 621-624.
 25. Giannopoulos L, Papaparaskevas J, Refene E. Mst typing of antimicrobial-resistant *Propionibacterium acnes* isolates from patients with moderate to severe acne vulgaris. *Anaerobe* 2015; 31: 50-54.
 26. Oprica C, Emtestam L, Lapins J. Antibiotic-resistant *Propionibacterium acnes* on the skin of

- patients with moderate to severe acne in stockholm. *Anaerobe* 2004; 10: 155-164.
27. Doğan B, Bektöre B, Karabacak E, Özyurt M. Resistance status of antibiotics in Gram-positive bacteria isolated from acne lesions in İstanbul. *Turkderm-Turk Arch Dermatol Venereology* 2017;51:32-6
 28. Nast A, Dreno B, Bettoli V. European evidence-based (S3) guidelines for the treatment of acne. *J Eur Acad Dermatol Venereol* 2017; 26 Suppl 1: 1-29.
 29. Dumont-Wallon G, Moyse D, Blouin E. Bacterial resistance in French acne patients. *Int J Dermatol* 2010; 49: 283-288.
 30. Nord CE, Oprica C. Antibiotic resistance in *Propionibacterium acnes*: Microbiological and clinical aspects. *Anaerobe* 2006; 12: 207-210.